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PATENT APPLICATION OF

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ENTITLED

COMPOSITION AND METHODS FOR STIMULATING
WOUND HEALING AND FIBROBLAST PROLIFERATION

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COMPOSITIONS AND METHODS FOR STIMULATING WOUND HEALING AND FIBROBLAST PROLIFERATION

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BACKGROUND OF THE INVENTION

a This invention relates to compositions and
5 methods to stimulate wound healing. More
particularly, this invention relates to the use of
compositions including placental alkaline phosphates
for stimulating wound healing.

10 People afflicted with long-term illness run
the risk of getting bed sores, pressure sores and a
myriad of skin irritations and chronic wounds.
Cancer patients, in particular breast cancer
patients, treated with radiation face the risk of
skin burns. Wound healing after surgical
15 intervention has been historically problematic. The
benefits of surgery, even in life threatening
situations, are offset by the formation of
disfiguring scar tissue. Adult wound healing is
characterized by fibrosis, scarring, and sometimes by
20 contracture.

Fibroblast cells, located in the dermal
layer, play important roles in wound healing by, for
example, producing components of the extracellular
matrix like collagen and various cytokines, which, in
25 turn, enhance the proliferation and migration of
keratinocytes. Keratinocytes are located in the
epidermal layer and form a barrier against the
external environment.

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Healing of wounds is the result of
interplay among different cell types and various
growth factors. Some of the growth factors,
including platelet-derived growth factor (PDGF),
5 epidermal growth factor (EGF), fibroblast growth
factor (FGF), transforming growth factor α (TGF- α),
and insulin-like growth factor I (IGF-I), are
considered to play significant roles by enhancing
proliferation of fibroblasts and/or keratinocytes
10 while TGF- β appears to primarily act via increasing
matrix formation.

Normal wound healing is also characterized
by the production of collagen. In adult skin,
fibroblasts present at the wound site usually produce
15 more collagen than necessary for optimal healing due
to the stimulatory actions of TGF- β and PDGF.
Excess collagen then leads to the undesirable
formation of scar tissue.

SUMMARY OF THE INVENTION

20 In a first aspect, the invention pertains to
a composition for wound healing in a patient comprising
placental alkaline phosphatase and a gel-forming
material.

In another aspect, the invention pertains to
25 a composition for stimulating cell proliferation
comprising placental alkaline phosphatase and a
proliferation compound. In these embodiments, the
proliferation compound is selected from the group

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consisting of a growth factor, a growth promoting serum factor and mixtures thereof.

In further aspects, the invention pertains to a method for accelerating wound healing applying a composition to the wound, the composition comprising placental alkaline phosphatase.

In addition, the invention pertains to a method for stimulating proliferation of cells in a cell culture. The method includes contacting the cells with a composition comprising a cell culture medium, placental alkaline phosphatase and a proliferation compound selected from the group consisting of a growth factor and a growth promoting serum factor.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a picture of a coomassie stained gel with purified placental alkaline phosphatase (PALP).

Fig. 2 is a picture of microplate wells with human skin fibroblasts incubated with fetal calf serum and with and without PALP.

Fig. 3 is a plot of DNA synthesis in human skin fibroblasts in the presence and absence of serum. Responses with PALP, INS and/or calcium are shown.

Fig. 4 is a plot of the effect of PALP concentration on cell proliferation after 3 days of incubation.

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Fig. 5 is a plot of the effect of PALP concentration on cell proliferation after 6 days of incubation.

Fig. 6 is a plot of the effect of partially purified PALP, concentration of PALP and concentration of serum on cellular proliferation.

Fig. 7 is a plot of the effect of PALP and growth factors on CC39SK human skin fibroblasts.

Fig. 8 is a plot of the effect of PALP and growth factors on CC986SK human skin fibroblasts.

Fig. 9 is a plot of the effect of PALP and growth factors on CC1058SK human skin fibroblasts.

Fig. 10 is a plot of the effect of PALP and growth factors on CC974SK human skin fibroblasts.

Fig. 11 is a plot of the effect of PALP and growth factors on CC944SK human skin fibroblasts.

Fig. 12 is a plot of the effect of PALP and growth factors on CC966SK human skin fibroblasts.

Fig. 13 is a plot of the effect of PALP and various growth factors on the amount of collagen in cells and media.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

Beneficial compositions that include placental alkaline phosphatase (PALP) can stimulate viability and proliferation of fibroblasts. Particularly when the PALP compositions are used in vitro for cell culture applications, the compositions can also include one or more proliferation compounds, which can be growth factors or growth promoting serum

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factors. Proliferation compounds synergistically combine with PALP to stimulate adult fibroblast proliferation and are referred to below as serum/growth factors. Serum/growth factors are optional when the PALP composition is used for in vivo wound healing applications, although use of serum/growth factors may be preferred. PALP compositions, when applied to a wound, are expected to accelerate the viability and the proliferation of the fibroblasts at the wound site resulting in accelerated wound healing.

PALP is an evolutionary new member of the alkaline phosphatase group of enzymes that hydrolyze phosphate-containing compounds and are reviewed in McLomb et al., "Alkaline Phosphatases," (Plenum Press, New York, 1979) and Millan and Fishman, "Biology of human alkaline phosphatases with special reference to cancer," Critical Rev. Clin. Sci., 22: 1-39 (1995), both of which are incorporated herein by reference. PALP is synthesized by human and primate placentas. PALP can exhibit growth factor-like effects in fetal fibroblasts, such as human fetal fibroblasts and mouse embryo fibroblasts. See, for example, She et al. (2000) FEBS Letters, 469, p.163-167 and She et al. (2000) Cell. Signal. 12, p.659-665, both of which are incorporated herein by reference. In the present approaches, contrary to earlier work, it has been discovered that PALP, in combination with serum/growth factors, stimulate

proliferation of adult fibroblasts, in particular adult skin fibroblasts. Without the serum/growth factors, PALP evidently does not significantly stimulate adult fibroblasts proliferation.

5 Agents that enhance the viability and numbers of fibroblasts can exert promoting effects on wound healing. In addition, PALP can enhance the proliferation of fibroblasts without exerting a stimulatory effect on collagen synthesis. PALP
10 compositions may be used on a variety of patients for wound healing, and the expectation is that healing incorporating PALP compositions will cause less scarring than growth factors alone. The patient can be an animal, especially a mammal. Preferably, the
15 patient is human, canine, porcine, bovine or the like.

Both semi-purified placenta extracts and highly purified PALP preparations can be used as active components in compositions aimed at
20 accelerating proliferation of fibroblasts. In vivo, semi-purified placenta extracts enriched in PALP or highly purified PALP preparations may accelerate the viability and proliferation rate of fibroblasts at wound sites. Specifically, in the presence of natural
25 serum and/or added growth factors, PALP is expected to induce a synergistic effect on fibroblast growth resulting in accelerated wound healing without a corresponding stimulation of collagen formation.

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In addition, PALP-containing compositions can be useful for improving in vitro cell culturing of adult fibroblast cells. In vitro, the presence of growth factors or growth promoting serum factors, however, is necessary for accelerating the viability and proliferation rate of adult fibroblasts. Serum/growth factors, as referred to herein, include agents that enable cells to move from stationary phase to the proliferative phase. As shown in the Examples below, the combination of PALP and serum/growth factors result in a synergistic effect on adult fibroblast proliferation.

PALP compositions generally include PALP and a non-toxic carrier or vehicle. For wound healing applications, the preferred form of the PALP composition is a gel. For either in vivo or in vitro applications, PALP compositions may include additional serum/growth factors to increase the proliferation of the fibroblasts. In addition, PALP compositions can include one or more additives/enhancers, such as preservatives, buffers, moisture control compounds and/or antibiotics. PALP compositions generally include sufficient PALP to stimulate proliferation of fibroblasts for the particular application.

The PALP compositions preferably enhance viability and proliferation of skin fibroblasts. The fibroblasts can be postnatally derived skin fibroblasts and/or embryonic fibroblasts. The PALP

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composition generally do not stimulate proliferation of keratinocytes.

5 In wound healing, shortly after skin is wounded, various growth factors, including, for example, PDGF, TGF- β 1, and other growth factors can be detected in the wounded tissue. Collagen synthesis can also occur at a wound site. An excess of collagen synthesis, however, can result in the formation of undesirable scar tissue at a wound site.

10 Application of a PALP composition to the wounded tissue is expected to accelerate wound healing via stimulation of fibroblast growth. It has also been discovered that PALP can also decrease, rather than increase, collagen synthesis by fibroblasts in the

15 presence of one or more growth factors, for example, PDGF and/or TGF- β 1. PALP compositions may, thus, accelerate wound healing with less scar formation. In contrast, if PDGF is applied alone for wound healing, in addition to stimulating cell

20 proliferation, the PDGF also stimulates collagen production that should lead to enhanced scarring.

In order to accelerate wound healing, PALP compositions can be applied to a wound. The PALP compositions can be applied in a variety of ways

25 including, but not limited to, direct application to the wound. Since serum/growth factors can be detected at a wound site as part of the natural wound healing process, addition of exogenous, i.e., non-native, progression factors, either included in the

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PALP composition or as a separate composition, can be optional for wound healing.

The PALP compositions described herein may be used to stimulate healing of a variety of wounds.

5 PALP compositions can be applied to external wounds including, for example, those caused by surgery, cuts, bruises, burns, sores and the like. PALP compositions may also be applied to internal organs, for example, during surgery. PALP compositions may
10 also be beneficial for cosmetic applications to enhance wound healing with less scarring.

Generally, the use of a composition that accelerates wound healing is desirable, because it would result in (i) reduction of period of
15 discomfort, (ii) shorter stay in the hospital after major surgeries (resulting in major cost savings), (iii) decreased potential for infections, (iv) faster recovery after cosmetic surgery, and (v) overall reduction in sick leaves due to major wounds.

20 PALP Products

PALP products are PALP compositions formulated for specific applications. The PALP products described herein include PALP and may include other active ingredients, such as serum/
25 growth factors and other additives. The PALP products generally also include a non-toxic carrier or the like to facilitate use of the PALP product. The nature of the PALP product including the types and amounts of both active ingredients and inactive

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carriers depend on the intended use of the PALP product. The PALP product can be used for in vivo wound healing or in vitro cell culturing.

PALP products can include placental material containing PALP, a semi-purified PALP preparation, a highly purified PALP preparation, recombinant PALP and the like. Human PALP is a protein, specifically, an enzyme found in the human placenta. PALP is a member of the group of alkaline phosphatase enzymes that hydrolyze phosphate containing compounds at alkaline pH values. At neutral pH they are observed to hydrolyze phosphatidic acid and some other substrates.

Natural PALP is observed to be a dimer of two identical glycosylated polypeptide subunits. The primary source of PALP is a placenta, which synthesizes this enzyme in latter stages of pregnancy, i.e. during approximately the second and third trimester of pregnancy in humans. At term in humans, PALP becomes a major alkaline phosphatase in circulation. PALP has been cloned and overexpressed in several cell lines, as described in Millan and Fishman, "Biology of human alkaline phosphatases with special reference to cancer," Critical Rev. Clin. Sci. 22:1-39 (1995), incorporated herein by reference. Production of recombinant PALP has been difficult so far, although low yields have been obtained in Escherichia Coli (see Beck and Burtsch, "Expression of human placental alkaline phosphatase

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in *Escherichia coli*," Protein Expression and Purification 5:192-197 (1994), incorporated herein by reference) and yeast Pichia pastoris (see Heimo et al., "Human placental alkaline phosphatase:Expression in *Pichia pastoris*, purification and characterization of the enzyme," Protein Expression and Purification 12:85-92 (1998), incorporated herein by reference).

Semi-purified PALP preparations can be obtained, for example, from Sigma Chemical Co. St. Louis, MO. The semi-purified PALP can be used to form PALP products or can be used to obtain highly purified PALP. Highly purified PALP from human placenta can be obtained by successive chromatographic separations, as described further in Example 1 below.

The concentration of PALP in a PALP product can have various values and generally depends on the specific use. For in vivo wound healing applications, the concentration may also depend on the location and type of wound, and the individual on whom it is used. Generally, the concentration of PALP in a wound healing product can be at least about 0.001 mg/1 g product, preferably between about 0.01 and about 0.5mg/1g product and more preferably, between about 0.01 and about 0.1mg/1g product.

For in vitro cell culture applications, a PALP product can be included with a cell culture media or, alternatively, diluted into a cell culture media by addition to a culture. The concentration of

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PALP products may include one or more serum/growth factors. Growth factors that can be included in PALP products include, for example, platelet-derived growth factor (PDGF), epidermal growth factor (EGF), fibroblast growth factor (FGF), transforming growth factor α (TGF- α), insulin-like growth factor I (IGF-I), transforming growth factor- β 1 (TGF- β 1) and insulin.


25 The serum/growth factors may augment the PALP effects by activating the fibroblast cells from a stationary phase in a cell cycle, referred to as G₀ cycle, to a proliferative phase, referred to as G_{1-m} cycle. PALP products may also include any other

agent that can induce cells into the proliferative phase.

The composition may include one growth factor, preferably PDGF. Alternatively, the
5 composition may include two growth factors, preferably PDGF and insulin. More than two growth factors can be included.

The concentration of the serum/growth factors in a PALP product, if used, can vary
10 depending on the specific growth factor used, the specific use of the composition, the nature of the wound and the like. Generally, the concentration of the serum/growth factor in a PALP product can be between about 5 nanogram/milliliter (ng/ml) and about
15 1000ng/ml. Preferably, the concentration of the serum/growth factors in the PALP composition is between about 25ng/ml and about 250ng/ml, and more preferably, the concentration is between about 50ng/ml and about 150ng/ml.

20 To incorporate growth promoting serum factors, a PALP product may include whole serum, as a composite of growth factors. Serum can be from a variety of sources, for example, fetal calf serum (FCS). Serum is generally obtained during blood
25 fractionation and is known in the art. Specifically, serum generally is a blood product with cells and cell fragments removed. Serum may also include serum fractions that have undergone other purification steps while retaining a complex mixture of compounds.



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Serum is generally sterile or is sterilized prior to addition to the composition. In embodiments for proliferating fibroblasts in vitro, FCS is particularly preferable. For in vitro proliferation, 5 the final concentration of the serum in the cell culture can be between about 0.1 percent by volume and about 20 percent by volume. Preferably, the final concentration of the serum in the cell culture is between about 0.5 percent by volume and about 12 10 percent by volume, more preferably, between about 1 percent by volume and about 10 percent by volume.

One or more isolated serum factors may also be included in the PALP product. Serum factors, referred to herein, are components that are normally 15 found in the serum. Serum factors can include, for example, growth factors, and other growth promoting agents, such as lysophosphotidic acid (LPA). It should be noted that tests have indicated that PALP and LPA do not promote each other's effects on 20 fibroblast growth. Isolated serum factor/s can be included in the PALP products to enhance PALP effects. The concentration of the serum factors in the PALP products, if used, can vary dependent on the specific serum factor and the specific use of a PALP 25 product.

Additives/enhancers in PALP compositions may include various ingredients, for example, preservatives (such as parabenes, quaternary ammonium compounds, alcohol, phenols, essential oil and the

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like), buffers, antioxidants (such as vitamine E), antimicrobials, vitamins, and moisture control agents (such as glycerine, propylene glycol, and the like). Other potential additives include, for example,

5 analgesics, anesthetics, anti-acne agents, anti-dermatitis agents, anti-pruritic agents, anti-inflammatory agents, anti-hyperkeratolytic agents, antiperspirants, anti-psoriatic agents, anti-seborrheic agents, anti-aging agents, anti-wrinkle

10 agents, skin lightening agents, depigmenting agents, corticosteroids, additional tanning agents or hormones. These additives may have some desirability in wound healing formulations.

For wound healing applications, the PALP

15 products can also include pharmaceutically and/or cosmetically acceptable carriers or vehicles. Preferably, the carriers are non-toxic. A pharmaceutically acceptable carrier does not cause an adverse physical reaction upon administration and is

20 one in which PALP is sufficiently soluble to deliver a therapeutically effective amount of PALP. The therapeutically effective amount of PALP may vary based on the individual patient, the indication being treated and other criteria evident to one of ordinary

25 skill in the art. A therapeutically effective amount of PALP is one sufficient to induce repair, healing and restoration of the target tissue to the original strength.

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Similarly, a cosmetically acceptable carrier is one that does not cause an adverse physical reaction upon application to the wound site and one in which PALP is sufficiently soluble to deliver a cosmetically effective amount of PALP. The cosmetically effective amount of PALP may vary based on the individual patient, the indication being treated and other criteria evident to one of ordinary skill in the art. A cosmetically effective amount of PALP is one sufficient to induce repair, healing or augmentation of a target tissue with minimal scar tissue.

As used herein, "pharmaceutical" or "cosmetic" will be understood to encompass both human and animal pharmaceuticals or cosmetics. Carriers and vehicles can be included in the PALP products in order to obtain an appropriate consistency, for example, gels, lotions, cream, rinse and the like. These products are suitable as topical compositions for wound healing applications.

Suitable carriers generally include, for example, water, acetone, ethanol, ethylene glycol, propylene glycol, butane-1,3-diol, isopropyl myristate, isopropyl palmitate, mineral oil, mixtures thereof and the like. Buffered solutions can also serve as carriers. Methodology and components for formulation of cosmetic and pharmaceutical compositions are well known, and can be found, for example, in Remington's Pharmaceutical Sciences,

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Eighteenth Edition, A.R. Gennaro, Ed., Mack Publishing Co. Easton, PA 1990, incorporated herein by reference.

In embodiments for topical applications, the carrier may be in any form appropriate for topical application to the skin including, for example, solutions, colloidal dispersions, emulsions (oil-in-water or water-in-oil), suspensions, creams, lotions, gels, foams, mousses, sprays and the like. PALP compositions suitable for use in topical application may also include, for example, liposomal carriers suspended in a suitable base or vehicle. Any liquid, pharmaceutically acceptable vehicle in which PALP is at least minimally soluble is suitable for topical use in the present invention. Other preparations that may be suitable include application of the composition onto a polyvinyl alcohol sponge.

For wound healing applications, the PALP products are preferably a gel. In some suitable embodiments, the gel includes methyl cellulose, agar, agarose, gelatin, calcium alginate or combinations thereof to form the gel. For example, the PALP after sterile filtration can be incorporated into sterile 3% by weight methyl cellulose gel, 1% by weight agarose gel, 4% by weight gelatin gel or 1-3% by weight calcium alginate. Generally, the PALP solution is sterile filtered and added to the gel formers in a sterile room. Wounds can be dressed

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The PALP composition can be stored under aseptic conditions at 4°C. Since PALP is stable at even room temperature and since growth factors and other typical additives are stable for several weeks at 4°C, the formulations should remain fully active for at least about 3 weeks.

For cell culture applications, the nature of the carrier just has to be consistent with subsequent inclusion in a cell culture medium. Generally, for dilution into a cell culture medium, 25 the carrier is an aqueous solution, and can be sterilized water or a buffer. PALP can also be stored as a powder. The PALP, in whatever form obtained, can be directly added to a cell culture

The cell culture medium should be suitable for culturing adult fibroblast cells. Suitable cell culture media for culturing adult fibroblast cells are known in the art. In particular, commercial cell culture media for culturing adult fibroblast cells are available, as described in the Examples below.

PALP compositions can be made using any number of suitable techniques. In some embodiments, PALP and a carrier can be mixed together within a commercial mixer to form a solution, a suspension or the like. In cosmetic and/or pharmaceutical composition embodiments, methodologies for formulation are well known, and can be found, for example, in Remington's Pharmaceutical Sciences, Eighteenth Edition, A.R. Gennaro, Ed., Mack Publishing Co. Easton, PA 1990, incorporated herein by reference.

The PALP compositions can be additionally processed after synthesis for purification, sterilization and the like. Sterilization, for example, can be conducted by filter sterilization, irradiation and the like. Methods for conducting these steps are known in the art and can be found, for example, in Remington's Pharmaceutical Sciences, Eighteenth Edition, A.R. Gennaro, Ed., Mack Publishing Co. Easton, PA 1990, incorporated herein by reference.

Stimulation of Fibroblast Proliferation and Wound Healing

The PALP products/compositions described herein can be used for increasing the viability and proliferation of cells, particularly fibroblasts. The PALP compositions are intended to be used, in vivo, in patients to accelerate wound healing. Furthermore, the PALP composition may be used for wound healing to reduce the amount of scar tissue formed at a wound site in a patient. In addition, the PALP compositions can be used in vitro in fibroblast cell cultures.

In vitro, the PALP compositions can be used in the presence of one or more serum/growth factors to stimulate viability and proliferation of fibroblasts. Cells are contacted with the PALP compositions for a period of time in the presence of growth stimulating factors, such as serum and/or growth factors. The PALP compositions are added to

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the cell culture medium to provide the necessary contact with the cells to stimulate cell proliferation.

5 The order of combining the components of the cell culture medium with the PALP composition and the progression factors generally is not significant. For example, the PALP compositions can be added directly to the cell cultures containers such as cell culture plates or flasks. The serum may be included
10 in the PALP composition. Alternatively, the serum may be provided separately to the cell culture container before, during or after addition of the PALP composition. Alternatively, the PALP composition, the serum/growth factors or both can be
15 combined with the cell culture medium prior to addition to the cell culture container. A combination of these approaches can be used to obtain a desired PALP concentration and serum/growth factor concentration with the passage of time during the
20 cell culture process.

In in vitro embodiments, the cells are contacted with the PALP composition and serum/growth factors for selected amount of time to stimulate cellular proliferation. Preferably, the cells are
25 contacted with the PALP composition for at least about one day. In other embodiments, the cells are contacted with the PALP composition for at least about 3 days and, for example, between about 3 days and about 6 days. In other embodiments, the cell

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cultures are maintained continuously in the presence of the PALP composition. During the cell culturing process, the cells are maintained in suitable conditions, such as temperature and oxygenation, to
5 provide appropriate conditions for cell viability and proliferation. Appropriate cell culturing conditions are well known in the art.

To promote wound healing, the PALP composition, or PALP alone, can be applied to a wound
10 either directly or indirectly. Routes of administration include, for example, topical, transdermal, parenteral, and gastrointestinal. Direct routes of applying PALP compositions to a wound are topical and involve contacting the PALP composition
15 directly to the wound. Transdermal administration can be accomplished by application of a cream, rinse, gel and the like that are capable of allowing the PALP to penetrate the skin. Parenteral administration can include, for example, electrical or direct injection
20 such as intravenous, intramuscular, intraperitoneal or subcutaneous injection. Gastrointestinal approaches involve oral injection of a composition.

Topical administration can be accomplished via a topically applied cream, rinse, gel and the
25 like that include PALP in the composition. In some embodiments, the creams, ointments and the like containing PALP can be delivered by dressings, bandages or other similar coverings capable of releasing a therapeutic amount of PALP. Such

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dressings can be directly placed on the wound to promote healing. Other methods of delivering the PALP compositions are also within the scope of this invention. The therapeutic amount of PALP that is
5 necessary to be delivered to the wound site can vary and depend on the specific wound, the location of the wound, the patient and the like.

The PALP composition can be administered to the wound for a suitable amount of period. Suitable
10 period is generally for at least one day. Suitable period can be for at least about 3 days. In some embodiments, PALP compositions may be administered to a wound for at least about a week. In other embodiments involving chronic problems, the PALP
15 compositions may be administered for several months or more.

The PALP composition can be administered as needed. Alternatively, PALP compositions can be administered two or more times a day. The frequency
20 of administration of the PALP composition can vary and depend on the type of wound, the location of the wound, the concentration of the PALP in the composition and the method used to administer the PALP composition. Generally, a therapeutically
25 effective amount of PALP composition is administered. A medical professional supervising treatment can adjust administered doses to obtain desired results.

Storage, Packaging and Distribution

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The PALP products can be stored, preferably at about room temperature if they do not contain other serum/growth factors or preferably at about 4° C if they do contain another serum/growth factor.

5 Preferred storage techniques minimize the risk of contamination and maintain the stability of the product. Following a final sterilization step, the sterile PALP product is transferred only using sterile transfer techniques, or the PALP product is
10 sterilized in the container that is sealed for sterile storage and/or transportation of the PALP product for distribution. For cell culture applications, maintaining sterile conditions is also important.

15 Appropriate packaging generally depends on the intended use of the PALP product. For cell culture applications, the PALP product generally is stored in a container for easy administration to a cell culture system in the form desired. For wound
20 healing applications, the packaging generally depends on the form of the product. Ointments and the like can be stored in tubes or the like. Liquid solutions can be stored in sealed vials. Capsules or the like for oral injections can be stored in sterile jars or
25 the like.

For distribution, the PALP composition can be placed in a sterile, sealed container. The containers are generally dated such that the date reflects the maximum advisable storage time

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Example 1- Purification of Human PALP

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A 350 mg quantity of commercial PALP was dissolved into 10 ml of buffer A (0.1 M sodium acetate, 0.5 M NaCl, 1 mM MgCl₂, 1 mM CaCl₂ at pH 6.5). The partially purified human PALP preparation

was further purified first by successive Concanavalin A-Sepharose and Q-Sepharose column chromatography. The Concanavalin A-Sepharose column was run using buffer A as the solvent. For elution, buffer A

5 included 50 mM α -methyl-D-mannopyranoside. The active fractions were pooled and dialyzed against buffer B (50 mM tris-HCL at pH 7.7). After dialysis, the protein was passed through a Q-sepharose column. The protein was eluted with buffer B using a linear

10 gradient of 0-250 mM potassium phosphate at a pH of 7.5. The active fractions from the Q-sepharose column were pooled and dialyzed against phosphate-buffered saline and concentrated by Amicon ultrafiltration.

15 Then, the purified PALP fraction off the Q-Sepharose column (which still contained 2 major proteins) was purified to homogeneity by t-butyl hydrophobic interaction chromatography (HIC). Prior to adding the protein to the t-butyl HIC column, the

20 solution was made 2M in ammonium sulphate, and the pH was adjusted to 6.8. The 5 ml bed volume t-butyl HIC cartridge (Bio-Rad) was connected to a FPLC system (Pharmacia). The column was eluted with buffer C (100 mM sodium phosphate buffer, 2M ammonium sulphate at

25 pH 6.8). The column was eluted with buffer C until a first protein peak completely eluted, and then a negative gradient of 2M-0M ammonium sulphate in 100 mM sodium phosphate at pH 6.8 was passed over the column. The negative linear gradient was used to

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elute a second enzymatically active peak. Following dialysis against phosphate buffered saline and concentration, purity of the enzyme was confirmed by SDS PAGE gel electrophoresis. A single protein band
5 was observed with an approximate molecular weight of 55-60 kD. The gel following separation was stained using coomassie blue for visual observation of protein bands. The purification procedure is described further in Chang et al. (1992) Eur. J.
10 Biochem. 209, 241-247, incorporated herein by reference.

PALP enzyme activity was assayed spectrophotometrically by monitoring the hydrolysis of 4-nitrophenylphosphate (as an increase in
15 absorbance at 410 nm) at room temperature (22°C). The analysis was performed with 1 ml volume containing 50 mM Na₂CO₃/NaHCO₃, 10 mM MgCl₂, 10 mM 4-nitrophenylphosphate at pH 9.8. The assay is described further in Chang et al. (1990) Biochem. J.
20 Vol. 272, pp.683-690, incorporated herein by reference. The extinction coefficient of 4-nitrophenol was taken as $1.62 \times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$. An enzyme activity of 1 U (unit) is defined as 1 μmol substrate hydrolyzed/min at 22°C at pH 9.8.

25 Fig. 1 shows a picture of a coomassie stained gel. The gel includes partially purified Sigma PALP preparation (shown in lane 2) further purified by successive Concanavalin A-Sepharose (lane 3), Q-Sepharose (lane 4), and t-butyl HIC

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chromatography (2 M - 0 M ammonium sulphate gradient)
(lane 5). Lane 1 shows the molecular mass standards.
Separation of proteins was performed by conventional
sodium dodecyl sulfate-polyethylene amine gel
5 electrophoresis (SDS-PAGE) and proteins were stained
by coomassie blue stain.

Figure 1 demonstrates that while the Sigma
preparation contains 3 major proteins (one of them is
PALP while an approximately 52 kDa protein band is α_1 -
10 antitrypsin) and several minor proteins, the purified
preparation apparently contains only PALP.
Identification of the PALP band by sequence analysis
was performed by the Mayo Clinic Protein Core
Facility (Rochester, MN, USA). Although the purified
15 PALP preparation did not seem to contain any other
protein, the possibility of minor protein
contaminants cannot be ruled out. But this
possibility does not diminish the usefulness of the
purified PALP.

20 EXAMPLES SHOWING THE EFFECTS OF PALP ON HUMAN CELL
LINES

In the following examples, the effects
of PALP and other factors on human cell lines are
examined. The following materials were used in these
25 examples.

Partially purified human PALP was purchased
and purified as described in Example 1. (3-4,5-
dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
(MTT) was purchased from Sigma Chemical Co., St.

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Louis, MO. Insulin (INS) and all the other growth factors used here were purchased from Boehringer Mannheim, Indianapolis, IN. [Methyl-³H]thymidine (500 mCi/mmol) was purchased from Dupont NEN, Boston, MA.

- 5 Tissue culture reagents, including Dulbecco's modified Eagle's medium (DMEM) and fetal calf serum (FCS) were purchased from GIBCO-BRL (Rockville, Maryland).

- 10 Human skin fibroblast cell lines were purchased from American Type Culture Collection. The cell lines, CCD 39 SK, CCD 986 SK, CCD 1058 SK, CCD 974 SK, CCD 944 SK, and CCD 966 SK, were derived respectively from one week old, 22 year old, 52 year old, 61 year old, 66 year old, and 78 year old
15 subjects. Each cell line, maintained in 10% FCS-containing DMEM, was used between 4-7 passages after thawing.

Example 2-Effect of PALP on Proliferation of Skin Fibroblasts and DNA synthesis

- 20 This example relates to the effect of PALP on a skin fibroblast culture and on the amount of DNA synthesis in the fibroblast culture.

- CCD 1058 SK human skin fibroblasts were seeded at 3×10^4 cells/well (12-well microplate) and
25 then incubated in 10% FCS-containing DMEM for 4 days in the absence (upper panel) or presence (lower panel) of 2 Unit/ml purified PALP. The visual results are shown in Fig. 2. Visual inspection of this and similar pictures clearly gave the impression

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that PALP treatment increased the number of fibroblasts. Initial visual analysis was followed by collection of quantitative data, including measurement of DNA synthesis and cell proliferation, as described in the following examples.

Example 3 - DNA Synthesis

This example describes the increased DNA synthesis from fibroblast cells resulting from PALP use.

10 CCD 1058 SK human skin fibroblasts were serum-starved (27 h) and then incubated for 18 h in the absence of serum (A) or in the presence of 2% FCS (B). During the incubation, the fibroblasts were untreated or treated with 1 Unit/ml purified PALP, 15 500 nanomolar (nM) insulin (INS) and 3.8 millimolar (mM) total Ca^{2+} alone or in combinations as indicated.

For the determination of DNA synthesis, human skin fibroblasts were grown in 12-well tissue culture dishes to about 40-50% confluency in 10% fetal calf serum (FCS)-containing DMEM, washed, and 20 then incubated in serum-free medium for 27 h. The cells were washed and then treated for 17 h in serum-free medium with PALP in the absence or presence of INS and/or calcium (Ca^{2+}), followed by incubation for 25 1 h in the presence of [^3H]thymidine (1 $\mu\text{Ci}/\text{well}$). At this point, cells in each well were sub-confluent. In these experiments, PALP was added to the cells 10 and 20 min prior to Ca^{2+} and INS, respectively. However, PALP was similarly effective when it was added 20 min

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prior to or 10 min after insulin. The cells were washed twice with PBS, then four times with 5% trichloroacetic acid, and finally twice with absolute ethanol. The acid-insoluble material, which contained
5 [3H]thymidine-labeled DNA, was redissolved in 0.3 M sodium hydroxide and then ³H activity was counted. Using this procedure, all [3H] activity is associated with DNA.

Fig. 3 shows the results of DNA synthesis
10 in fibroblasts. Data are the means \pm S.E.M. (standard error of means) of three experiments each performed in triplicate. On the plot shown in Fig. 3, the asterisks indicate the following:
*significantly ($P < 0.01$ level) greater values than the
15 value with no addition of additional factors;
*^asignificantly ($P < 0.01$ level) greater values than obtained with PALP or INS alone; *^bsignificantly ($P < 0.01$ level) greater values than the combined effects of Ca^{2+} and INS. The last value is the same
20 as the effect of Ca^{+2} alone, which are not shown.

The data indicate that in the absence of serum, the presence of both high concentration of Ca^{2+} and INS is required for the stimulatory effect of PALP on DNA synthesis, while in the presence of
25 serum, PALP also had stimulatory effect in the absence of Ca^{2+} and INS. Cell viability/proliferation was measured directly using MTT analysis, as described in the following example.

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EXAMPLES 4-12 - CELL VIABILITY/PROLIFERATION
MEASUREMENTS

These examples examine the quantitative effects on cell proliferation as a result of adding
5 PALP with or without other factors to cell cultures of human fibroblasts.

(MTT Assay) For the determination of Cell Viability/Proliferation, fibroblasts were seeded at 1000 cells/well in 96-microwell plates in 10% FCS-
10 containing DMEM. After 24 h, the medium was replaced with 1, 2, or 10% serum-containing fresh medium followed by treatments with PALP and growth factors for 3 or 6 days; in the latter case, after 3 days of treatments, the medium was changed for fresh
15 (respective) medium followed by re-treatment of cells. At the time of analysis, cells in each well were subconfluent. The relative changes in the number of viable cells were determined by MTT assay. This colorimetric assay is based on the ability of healthy
20 cells (mostly the mitochondrial compartment) to reduce MTT to a blue formazan product. A Labsystem Multiskan MS microplate reader purchased from Labsystemes (Franklin, MA) was used to measure the formation of formazan as an increase in absorbance at
25 a test wavelength of 540 nm and a reference wavelength of 690 nm. This technique is a widely used and accepted method to accurately determine the relative numbers of viable cells. In most cases, when the test agent does not strongly influence the

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oxidation-reduction balance of cells, the MTT assay is essentially a proliferation assay. The MTT assay is described further in Carmichael et al. (1987) Cancer Res. 47, p.936-942, incorporated herein by
5 reference.

In the data analysis, data were either mean values \pm one standard deviation (S.D.) (3-8 independent incubations from the same experiment; for evaluating significance, Student's *t* test is used) or
10 mean values \pm S.E.M. of three experiments (3 incubations in each; for evaluating significance, unpaired *t* test is used). Each experiment presented here was repeated at least once with similar results.

Example 4 - Combination of PALP and Insulin

15 This example shows the effect of combining purified PALP and insulin on fibroblast cell proliferation after either 3 days of incubation and after 6 days of incubation.

In a first set of experiments, CCD 1058 SK
20 human skin fibroblasts cells, incubated in 2% FCS-containing DMEM for 3 days, were treated with 0-2 Unit/ml purified PALP, as indicated, in the absence (●) or presence (▲) of 500 nM INS during the entire incubation period. The incubation period was followed
25 by an MTT assay. Fig. 4 shows a plot of MTT assay results. Data are means \pm S.D. of 8 incubations. As indicated in Fig. 4 with an "*", significantly ($P < 0.01$ level) greater values were obtained with the addition of PALP in comparison with the corresponding values

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in the absence of PALP. This experiment showed that 0.5-2.0 Unit/ml PALP increased the number of CCD 1058 SK fibroblasts both in the absence and presence of INS during a 3 day incubation period. However, PALP and insulin had greater effects in combination than alone.

In another experiment, CCD 1058 SK human skin fibroblasts cells, incubated in 2% FCS-containing DMEM for 6 days, were treated with 0-2 Unit/ml purified PALP, as indicated, in the absence (●) or presence (▲) of 500 nM INS during the entire incubation period. This was followed by the MTT assay. Fig. 5 shows a plot of the assay results. Data are means \pm S.D. of 8 incubations. As noted in Fig. 5 with an "*", significantly ($P < 0.01$ level) different values were obtained for the samples with added PALP from the corresponding values in the absence of PALP.

This experiment showed that 0.5-2.0 Unit/ml PALP increased the number of CCD 1058 SK fibroblasts both in the absence and presence of INS even if the incubation period was extended to 6 days. In the presence of 2% FCS, PALP and INS had no cumulative or synergistic effects on cell proliferation.

In the total absence of serum, PALP did not enhance proliferation of CCD 1058 SK fibroblasts (not shown). In contrast, in mouse embryo fibroblasts (NIH 3T3), PALP stimulated DNA synthesis and cell proliferation even in the absence of serum. In further experiments, in the absence of serum, PALP

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did not enhance cell proliferation in any of the other human skin fibroblast lines examined in this application.

Example 5 - Partly Purified PALP and Insulin

5 This example demonstrates that even partly purified PALP can be effective to stimulate fibroblasts in cell culture when combined with other agents.

10 CCD 1058 SK human skin fibroblast cells were treated for 6 days with no PALP (\square), or with 0.5 Unit/ml Sigma PALP (\boxtimes), or with 2 Unit/ml Sigma PALP (\blacksquare) in the presence of 2% or 10% FCS with or without 500 nM INS, as indicated. This was followed by the MTT assay. Fig. 6 shows a plot of the assay results.

15 Data were the means \pm S.D. of 8 incubations. As indicated in Fig. 6 with an "*", significantly ($P < 0.01-0.05$ level) greater proliferation was measured in sample with PALP than the corresponding values measured in the absence of Sigma PALP.

20 This experiment demonstrated that partially purified PALP also increases proliferation of CCD 1058 SK fibroblasts in the presence of 2-10% serum \pm INS. Therefore, for the same effect, a partially purified PALP preparation may be used instead of

25 highly purified PALP.

Example 6 - Effect of PALP Combined with Different Factors in One Human Skin Fibroblast Cell Line.

 This example relates to determining the effect of PALP, alone and in combination with other

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factors, on the viability and proliferation of the CCD 39 SK skin fibroblast cell line.

CCD 39 SK human skin fibroblasts, incubated in 1% FCS-containing DMEM for 6 days, were untreated (□) or treated (■) with 0.5 Unit/ml Sigma PALP in the absence or presence of PDGF (50 ng/ml), epidermal growth factor (EGF) (50 ng/ml), basic fibroblast growth factor (bFGF) (20 ng/ml), transforming growth factor- α (TGF- α) (50 ng/ml), TGF- β 1 (5 ng/ml), insulin-like growth factor I (IGF-I) (50 ng/ml) or INS (500 nM), as indicated. The incubation was followed by an MTT assay.

Fig. 7 shows a plot of the assay results. At these concentrations, the growth factors had maximal effects on proliferation in CCD 1058 SK fibroblasts, as determined in previous experiments (not shown). Data are the means \pm S.D. of 8 incubations. As indicated in Fig. 7 with an "*", significantly ($P < 0.01-0.05$ levels) greater cell proliferation was observed in the experiments in the presence of the PALP than in the corresponding experiments in the absence of PALP.

In these cells, PDGF was about twice as effective as PALP in increasing cell proliferation. However, PALP further increased the effect of PDGF, it reversed the inhibitory influence of TGF- β 1, and it increased the effects of IGF-I and INS. Overall, PALP had only positive effects on cell proliferation.

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Example 7 - Effect of PALP with Different Factors in
a Second Cell Line

This example relates to determining the effect of PALP, alone and in combination with other
5 factors, on the viability and proliferation of the CCD 986 SK skin fibroblast cell line.

CCD 986 SK human skin fibroblast cells, incubated in 1% FCS-containing DMEM for 6 days, were untreated (\square) or treated (\blacksquare) with 0.5 Unit/ml Sigma
10 PALP in the absence or presence of PDGF (50 ng/ml), EGF (50 ng/ml), bFGF (20 ng/ml), TGF- α (50 ng/ml), TGF- β 1 (5 ng/ml), IGF-I (50 ng/ml) or INS (500 nM), as indicated. This was followed by the MTT assay. Fig. 8 shows a plot of the assay results. Data are the
15 means \pm S.D. of 8 incubations. As indicated in Fig. 8 with an "*", significantly ($P < 0.01-0.05$ levels) greater assay values were obtained in the presence of PLAP than the corresponding values in the absence of PALP.

20 In this cell line, PALP, PDGF, bFGF, and TGF- α were similarly effective stimulators of cell proliferation while TGF- β 1 was clearly inhibitory. PALP enhanced the stimulatory effects of PDGF, it retained its stimulatory effects in the presence of
25 IGF-I and INS, and it reversed the inhibitory influence of TGF- β 1. Overall, PALP had only positive effects on cell proliferation in this cell line.

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Example 8 - Effect of PALP with Different Factors in
a Third Cell Line

This example relates to determining the effect of PALP, alone and in combination with other
5 factors, on the viability and proliferation of the CCD 1058 SK skin fibroblast cell line.

CCD 1058 SK human skin fibroblast cells, incubated in 1% FCS-containing DMEM for 6 days, were untreated (\square) or treated (\blacksquare) with 0.5 Unit/ml Sigma
10 PALP in the absence or presence of PDGF (50 ng/ml), EGF (50 ng/ml), bFGF (20 ng/ml), TGF- α (50 ng/ml), TGF- β 1 (5 ng/ml), IGF-I (50 ng/ml) or INS (500 nM), as indicated. This was followed by the MTT assay. Fig. 9 shows a plot of the assay results. Data are the
15 means \pm S.D. of 8 incubations. As indicated in Fig. 9 with an "*", significantly ($P < 0.01-0.05$ levels) greater values were obtained with PALP present than the corresponding values in the absence of PALP.

In this cell line, PALP was about half as
20 effective as PDGF but as effective as bFGF and TGF- α in enhancing cell proliferation. PALP enhanced the stimulatory effects of PDGF, bFGF, TGF- α , IGF-I, and INS and it effectively reversed the slight inhibitory influence of TGF- β 1. Overall, PALP had only positive
25 effects on cell proliferation in this cell line.

Example 9 - Effect of PALP with Different Factors in
a Fourth Cell Line

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This example relates to determining the effect of PALP, alone and in combination with other factors, on the viability and proliferation of the CCD 974 SK skin fibroblast cell line.

5 CCD 974 SK human skin fibroblasts cells, incubated in 1% FCS-containing DMEM for 6 days, were untreated (□) or treated (■) with 0.5 Unit/ml Sigma PALP in the absence or presence of PDGF (50 ng/ml), EGF (50 ng/ml), bFGF (20 ng/ml), TGF- α (50 ng/ml),
10 TGF- β 1 (5 ng/ml), IGF-I (50 ng/ml) or INS (500 nM), as indicated. This was followed by the MTT assay. Fig. 10 shows a plot of the assay results. Data are the means \pm S.D. of 8 incubations. As indicated in Fig. 10 with an "*", significantly ($P < 0.01-0.05$ levels)
15 greater values were obtained in the presence of PALP than the corresponding value in the absence of PALP.

This cell line responded very well to PDGF but less well to PALP and the other growth factors. PALP partially reversed the slight inhibitory
20 influence of TGF- β 1, it slightly enhanced the stimulatory effects of IGF-I and INS, and it retained its stimulatory effect in the presence of EGF and TGF- α . Overall, PALP had only positive effects on cell proliferation in this cell line.

25 Example 10 - Effect of PALP with Different Factors in a Fifth Cell Line

This example relates to determining the effect of PALP, alone and in combination with other

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factors, on the viability and proliferation of the CCD 944 SK skin fibroblast cell line.

CCD 944 SK human skin fibroblast cells, incubated in 1% FCS-containing DMEM for 6 days, were untreated (\square) or treated (\blacksquare) with 0.5 Unit/ml Sigma PALP in the absence or presence of PDGF (50 ng/ml), EGF (50 ng/ml), bFGF (20 ng/ml), TGF- α (50 ng/ml), TGF- β 1 (5 ng/ml), IGF-I (50 ng/ml) or INS (500 nM), as indicated. This was followed by the MTT assay. Fig. 11 shows a plot of the assay results. Data are the means \pm S.D. of 8 incubations. As shown in Fig. 11 with an "*", significantly ($P < 0.01-0.05$ levels) greater values were obtained in the presence of PALP than the corresponding value in the absence of PALP.

In this cell line, PDGF was a more effective inducer of cell proliferation than PALP, while PALP was as effective as bFGF and TGF- α . PALP enhanced the stimulatory effects of PDGF, bFGF, IGF-I, and INS, and it reversed the substantial inhibitory effect of TGF- β 1. Overall, PALP had only positive effects on cell proliferation in this cell line.

Example 11 - Effect of PALP with Different Factors in a Sixth Cell Line

This example relates to determining the effect of PALP, alone and in combination with other factors, on the viability and proliferation of the CCD 944 SK skin fibroblast cell line.

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CCD 966 SK human skin fibroblast cells, incubated in 1% FCS-containing DMEM for 6 days, were untreated (□) or treated (■) with 0.5 Unit/ml Sigma PALP in the absence or presence of PDGF (50 ng/ml), EGF (50 ng/ml), (bFGF) (20 ng/ml), TGF- α (50 ng/ml), TGF- β 1 (5 ng/ml), IGF-I (50 ng/ml) or INS (500 nM), as indicated. This was followed by the MTT assay. Fig. 12 shows a plot of the assay results. Data are the means \pm S.D. of 8 incubations. As indicated in Fig. 12 with an "*", significantly ($P < 0.01-0.05$ levels) greater values were obtained in the presence of PALP than the corresponding values in the absence of PALP.

In this cell line, PDGF was about twice as effective as PALP in enhancing cell proliferation, while PALP was more effective than the other growth factors. PALP increased the effects of each stimulatory growth factor and it also reversed the inhibitory influence of TGF- β 1. Overall, PALP had only positive effects on cell proliferation in this cell line.

Example 12 - Determination of Changes in Cell Number

This Example demonstrates that cell counting and MTT assays yield roughly equivalent results for determining the effectiveness of PALP in stimulating fibroblasts.

For the determination of changes in cell numbers, CCD 1058 SK fibroblasts were seeded in 12-well plates in 10% FCS-containing DMEM up to about ~20% confluency. Then, the medium was changed for

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fresh medium with no serum or supplemented with 2% or 10% FCS. Half of the wells received no further addition, while the other half of the cells were treated with 1 unit/ml PALP; both sets were incubated for 4 days. Cells were stained (Giemsa), then from each well 3 photographs were taken, and finally cells were counted manually.

In cell proliferation studies, after treatments for 4 days, in serum-free medium PALP had practically no effects, while in the presence of 2% and 10% FCS it enhanced cell numbers 39 and 33%, respectively. Parallel studies using the MTT viability assay (see above) showed that PALP had no effects in the absence of FCS, while in the presence of 2% and 10% FCS it increased the number of viable cells 44% and 30%, respectively. Since the experimental error was between 5-10%, it was concluded that in case of PALP, cell counting and the MTT assay provide practically the same results (i.e. suggesting that PALP does not alter the cellular oxidation/reduction events), i.e. that increased formazan production is proportional with increased cell numbers.

Example 13-Collagen Synthesis in the presence of PALP

This example relates to evaluating the amount of collagen synthesis in skin fibroblasts in the presence of PALP.

The method used here for determination of collagen synthesis is widely used and well accepted

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in the art. Fibroblasts at 80% confluency were incubated (in 6-well plates) in 1% FCS-containing DMEM for 48 h with [2,3-³H]proline (4 μ Ci/well), ascorbic acid (50 μ g/ml) and β -aminopropionitrile (50 μ g/ml); other agents present are indicated in Fig. 13. After incubations, the medium was harvested and the cells were washed with phosphate buffered saline and trypsinized in 0.5 ml 50 mM Tris-HCl, pH 7.6. Cells were sonicated and then incubated with ribonuclease (5 μ g/sample; 10 min). Proteins from both the cells and medium were precipitated with 10% trichloroacetic acid (TCA). The precipitate was washed with 5% TCA, and the pellet was dissolved in 0.5 ml 0.2 M NaOH. The samples were neutralized with 0.1 ml Hepes buffer, 120 mM (pH 7.2, containing 12.5 mM N-ethyl maleimide and 2.5 mM CaCl₂), and 0.16 ml of 0.15 M HCl, followed by collagenase (20 units/ml) treatment for 90 min at 37°C. Then, proteins were precipitated by an equal volume of TCA/tannic acid (10% and 0.5 %, respectively) and the supernatant was collected. The precipitate was washed again and the supernatants were combined. The counts from samples without collagenase treatment were determined and subtracted from that measured from collagenase-treated samples to estimate the amount of synthesized collagen.

To obtain collagen synthesis results for PALP treated fibroblasts, CCD 1058 SK cells were incubated in 1% FCS for about 48 hours. During the whole incubation period, the incubation medium

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contained [³H]proline and the cells were treated with 50 ng/ml PDGF, 5 ng/ml TGF- β 1, or 500 nM INS, as indicated, in the absence (p) or presence (v) of 1 Unit/ml Sigma PALP. The amount of [³H]collagen was
5 determined both in the cells (A) and in the medium (B). Data are the means \pm S.D. of 3 independent incubations. Fig. 13 shows a plot of the assay results.

Each growth factor shown, but not PALP,
10 considerably enhanced collagen synthesis, with 50% or more collagen being secreted into the medium. The growth factors did not appear to specifically stimulate collagen secretion. PALP alone had no stimulatory effects, while it moderately, but
15 invariably, reduced the effects of growth factors on both collagen synthesis and on the collagen content in the medium.

The embodiments above are intended to be
20 exemplary and not limiting. Additional embodiments are within the claims below. Although the present invention has been described with reference to preferred embodiments, workers skilled in the art will recognize that changes may be made in form and
25 detail without departing from the spirit and scope of the invention.

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